

Splice Modulation Therapy in Inherited Retinal Diseases

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Background: Antisense oligonucleotides (AONs) are small DNA/RNA molecules able to modulate pre-mRNA splicing, including the correction of aberrant splicing processes due to genetic mutations. Therapeutically, AONs are employed for a variety of inherited diseases, including Duchenne muscular dystrophy, spinal muscular atrophy, metabolic disorders etc. In this talk, I will mainly focus on the use of AONs for inherited retinal diseases (IRD), a group of disorders that is highly heterogeneous, both at the genetic and clinical level. Our proof-of-concept work on the correction of a recurrent splicing defect underlying *CEP290*-associated Leber congenital amaurosis (LCA), one of the most severe subtypes of IRD, has led to the initiation of a clinical trial, with positive interim results, demonstrating safety and in some subjects efficacy upon intra-ocular AON delivery. We expanded the use of AONs for the treatment of other subtypes of IRD, including Stargardt disease which is caused by *ABCA4* mutations.

Methods: Recently, we established a panel of *ABCA4* midgenes together encompassing the complete *ABCA4* gene, with which the effect of any *ABCA4* variant on pre-mRNA splicing can readily be assessed *in vitro*. Following site-directed mutagenesis, wild-type or mutant midgenes were transfected into HEK293T cells to identify variants affecting *ABCA4* pre-mRNA splicing. For variants resulting in aberrant *ABCA4* splicing, AONs were designed, and co-transfected with the midgenes to assess their efficacy to correct splicing defects. For several variants, patient-derived fibroblasts and/or iPSC-derived photoreceptor progenitor cells were treated with the AONs, and rescue efficacy was determined at the RNA level.

Results: In total, we identified more than a dozen different *ABCA4* variants that affect pre-mRNA splicing. The majority of variants resides in introns, and leads to the insertion of pseudoexons that are predicted to disrupt the reading frame and thus loss of *ABCA4* protein function. Another, recurrent, *ABCA4* variant (c.768G>T) led to an exon extension of 35 nucleotides(nt), thereby also disrupting the reading frame. Whereas some variants showed splicing defects in all cell types tested, others showed a more retina-specific splicing defect. For all variants that resulted in pseudoexon insertion, this process could be corrected by one or more AONs specifically targeting the corresponding pseudoexon. Intriguingly, also the 35-nt extension caused by the common c.768G>T variant could be rescued completely by blocking the alternative splice donor site in intron 6, forcing the spliceosome to use the original splice donor site of exon 6.

Conclusion: AONs appear to be an effective and versatile tool to correct different types of splice defects that are caused by *ABCA4* mutations. Given the promising data obtained so far in a clinical trial using AONs for the treatment of *CEP290*-associated LCA, AONs may serve as a broadly applicable therapeutic strategy, not only for Stargardt disease but potentially also for other genetic subtypes of inherited retinal disease caused by splicing defects.